Total Protein Normalization: Publications List

V3 Western Workflow[™]

Methodology Papers

Total Protein Loading Control vs. Housekeeping Protein Loading Control

Moritz CP et al. (2014).

Epicocconone staining: A powerful loading control for western blots. Proteomics14, 162–168.

Eaton SL et al. (2013).

Total protein analysis as a reliable loading control for quantitative fluorescent western blotting. PLoS One 8, e72457.

Gilda JE and Gomes AV (2013).

Stain-free total protein staining is a superior loading control to β -actin for western blots. Anal Biochem 440,186–188.

Colella AD et al. (2012).

Comparison of stain-free gels with traditional immunoblot loading control methodology. Anal Biochem 430, 108–110.

Lanoix D et al. (2012).

Stability of reference proteins in human placenta: general protein stains are the benchmark. Placenta 33, 151–156.

Suzuki O et al. (2011).

Use of sample mixtures for standard curve creation in quantitative western blots. J Exp Anim 60, 193–196.

Welinder C and Ekblad L (2011).

Coomassie staining as loading control in western blot analysis. J Proteome Res 10, 1416–1419.

Hagiwara M et al. (2010).

Application of SYPRO Ruby- and Flamingo-stained polyacrylamide gels to western blot analysis. Anal Biochem 397, 262–264.

Romero-Calvo I et al. (2010).

Reversible Ponceau staining as a loading control alternative to actin in western blots. Anal Biochem 401, 318–320.



Bulletin 6349

Aldridge GM et al. (2008).

The use of total protein stains as loading controls: an alternative to high-abundance single-protein controls in semi-quantitative immunoblotting. J Neurosci Methods 172, 250–254.

Selected Research Papers from JBC, PNAS, MCB, and JCS

Applying Total Protein Loading Control (e.g. Coomassie) in Western Blotting

Gene Regulation – Genetics, Epigenetics, Cell Biology, Neuroscience, Developmental Biology

Furuya F et al. (2013). Ligand-bound thyroid hormone receptor contributes to reprogramming of pancreatic acinar cells into insulin-producing cells. J Biol Chem 288, 16155–16166.

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The leucine-rich pentatricopeptide repeat-containing protein (LRPPRC) does not activate transcription in mammalian mitochondria. J Biol Chem 288, 15510-15519.

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Developmentally regulated linker histone H1c promotes heterochromatin condensation and mediates structural integrity of rod photoreceptors in mouse retina. J Biol Chem 288, 17895–17907.

Bryant JM et al. (2012).

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Cui CY et al. (2012).

Forkhead transcription factor FoxA1 regulates sweat secretion through Bestrophin 2 anion channel and Na-K-Cl cotransporter 1. Proc Natl Acad Sci USA 109, 1199–1203.

Gu S et al. (2012).

Basic helix-loop-helix transcription factor Twist1 inhibits transactivator function of master chondrogenic regulator Sox9. J Biol Chem 287, 21082–21092.

Song X et al. (2012).

Synergistic and multidimensional regulation of plasminogen activator inhibitor type 1 expression by transforming growth factor type β and epidermal growth factor. J Biol Chem 287,12520–12528.

Ahmady E et al. (2011).

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Intestinal hypoxia-inducible factor- 2α (HIF- 2α) is critical for efficient erythropoiesis. J Biol Chem 286,19533–19540.

Banerjee D et al. (2011).

Preferential repair of oxidized base damage in the transcribed genes of mammalian cells. J Biol Chem 286, 6006–6016.

Dixit A et al. (2011).

Dynamics of the T4 bacteriophage DNA packasome motor: endonuclease VII resolvase release of arrested Y-DNA substrates. J Biol Chem 286, 18878–18889.

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Signal Transduction - Neurobiology, Microbiology, Disease Mechanism, Cell Biology

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