

# MicroPulser™ electroporator





# A simple tool for transformation

The MicroPulser is a simple yet versatile electroporator that provides a safe and reproducible way for you to transform bacteria, yeast, and other microorganisms. Transformation efficiencies much higher than those obtained by chemical methods can be achieved by electroporation. A pulse is easily delivered by choosing a preset program and touching a single button.



#### **Unique Features of the System Include:**

- Faster sample handling Simple one-button pulse delivery, attached cuvette chamber, and rapid charge time
- Rapid program selection Preset, optimized programs for commonly studied bacteria and fungi
- Arc quenching (ARQ) system that reduces arcing, protecting against loss of valuable samples
- Broad range of parameters for manual optimization.
   Manual programming allows voltage to be selected in a 200–3,000 V range with 10 V precision, and pulse width to be adjusted in a 1.0–4.0 ms range with 0.1 ms precision
  - 3,000 V capability for improved efficiency in cuvettes with larger volume
    - Compact, space-saving design
    - Audible and visual pulse indicators
    - Display of time constant and actual voltage delivered to monitor reproducibility

#### **Programmed Functions**

	Program	Species	Cuvette Size	Preset Conditions
Bacteria	Ec1	Escherichia coli	0.1 cm	1.80 kV, 1 pulse
	Ec2	Escherichia coli	0.2 cm	2.50 kV, 1 pulse
	StA	Staphylococcus aureus	0.2 cm	2.50 kV, 1 pulse, 2.5 ms
	Agr	Agrobacterium tumefaciens	0.1 cm	2.20 kV, 1 pulse
	Ec3	Escherichia coli	0.2 cm	3.00 kV, 1 pulse
Fungi	Sc2	Saccharomyces cerevisiae	0.2 cm	1.50 kV, 1 pulse
	Sc4	Saccharomyces cerevisiae	0.2 cm	3.00 kV, 1 pulse
	ShS	Schizosaccharomyces pombe	0.2 cm	2.00 kV, 1 pulse
	Dic	Dictyostelium discoideum	0.4 cm	1.00 kV, 1 pulse, 1.0 ms
	Pic	Pichia pastoris	0.2 cm	2.00 kV, 1 pulse

Unless the pulse time is truncated below 5 ms, the unit will deliver the optimal time constant of ~5 ms to samples in high-resistance media.



## Why Electroporation?

#### **Efficient**

Electroporation is the most efficient transformation method available. It is orders of magnitude more efficient than chemical methods and provides more reproducible results than any other method. The MicroPulser is designed to deliver optimum electrical conditions for electroporation of *E. coli*, fungi, and other microorganisms, resulting in the highest efficiencies possible. The preset conditions are optimized for common bacteria and fungi. Voltage and pulse time can also be set manually, enabling you to optimize transformation conditions for your experiment.

#### **Compact and User-Friendly**

The all-in-one design and preset conditions precisely deliver the optimal parameters for bacteria and fungi, established by Bio-Rad and verified in the literature over the years. This simple optimization allows efficient transformation, with a minimum of effort, in practically no time! The small footprint saves valuable benchspace.

#### **Flexible**

You can choose voltages between 200 and 3,000 V, to transform the widest range of microorganisms. By using a larger-capacity cuvette and increasing voltage to maintain the same field strength, you can process large samples and increase your throughput.

### Cuvettes

Reproducible electroporation results require high-quality electroporation cuvettes for consistent pulse delivery to your valuable sample.

#### **Bio-Rad Cuvette Features:**

- High-quality construction for consistent performance
- Gamma-irradiated to ensure sterility
- Color-coded caps for easy identification
- Available in various package sizes

#### **Example Results**

Species	Strain	Volume	Efficiency
E. coli	DH10B	20 µl	1.6 x 10 <sup>10</sup>
E. coli	DH10B	20 μΙ	3.2 x 10 <sup>9</sup>
S. aureus	RN4220	50 µl	1.2 x 10⁵
A. tumefaciens	LBA4404	20 µl	7.0 x 10 <sup>6</sup>
E. coli	DH10B	20 µl	9.1 x 10°
S. cerevisiae	Sc948	40 µl	8.1 x 10 <sup>4</sup>
S. cerevisiae	Sc948	80 µl	2.2 x 10⁵
S. pombe	CHP408	200 μΙ	1.4 x 10 <sup>4</sup>
D. discoideum	KAx3	800 µl	88
P. pastoris	X33	40 µl	1.6 x 10⁴
	E. coli E. coli S. aureus A. tumefaciens E. coli S. cerevisiae S. cerevisiae S. pombe D. discoideum	E. coli         DH10B           E. coli         DH10B           S. aureus         RN4220           A. tumefaciens         LBA4404           E. coli         DH10B           S. cerevisiae         Sc948           S. cerevisiae         Sc948           S. pombe         CHP408           D. discoideum         KAX3	E. coli         DH10B         20 μl           E. coli         DH10B         20 μl           S. aureus         RN4220         50 μl           A. tumefaciens         LBA4404         20 μl           E. coli         DH10B         20 μl           S. cerevisiae         Sc948         40 μl           S. cerevisiae         Sc948         80 μl           S. pombe         CHP408         200 μl           D. discoideum         KAX3         800 μl

Experiments were carried out in an ordinary laboratory and represent average efficiencies obtained using the preset programs. The efficiencies were measured per 1 µg plasmid by stable expression using auxotrophic mutants or antibiotic resistance, with the exception of *D. discoideum*, which had efficiency measured using transient expression of GFP.

#### References

Becker DM and Guarente L, High-efficiency transformation of yeast by electroporation, Methods Enzymol 194, 182–87 (1991)

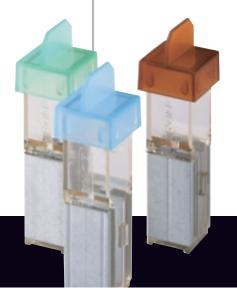
Cregg JM and Russell KA, Transformation, Methods Mol Biol 103, 27–39 (1998)

Howard PK et al., Establishment of a transient expression system for Dictyostelium discoideum, Nucleic Acids Res 16, 2613–2623 (1988)

Nickoloff JA (ed.), Electroporation protocols for microorganisms, Methods Mol Biol 47 (1995)

Prentice HL, High efficiency transformation of *Schizosaccharomyces pombe* by electroporation, Nucleic Acids Res 20, 621 (1992)

Sambrook J et al., Molecular Cloning: A Laboratory Manual, 2nd edition, Cold Spring Harbor Laboratory Press, Plainview, NY (1989)



#### **MicroPulser System Specifications**

Input voltage In-line switching, 100-120 V or

220-240 V

Input current 2 amp RMS (100-120 V)

1 amp RMS (220-240 V)

Maximum output voltage and current 3,000 V peak into > 600  $\Omega$ 

Limited to 100 A peak maximum

Decaying or truncated exponential waveform with RC time constant of

5.0 ms assuming loads of 3.3  $\mbox{k}\Omega$ 

200-3,000 V range with Output voltage adjustment

10 V adjustment

Pulse-time adjustment In manual mode, set time range is 1.0-4.0 ms with 0.1 ms precision

(providing the sample-determined pulse width has time constant >4.0 ms)

Operating environment Temperature 0-35°C

Humidity 0-95% without condensation

Regulatory

Meets EN61000-3-2 and EN61000-3-3

harmonic, flicker, and voltage fluctuation standards, FCC, Class A

IEC 1010, CE

Dimensions 31 x 21 x 8 cm (L x W x H)

Weight 2.9 kg

#### **Ordering Information**

Output waveform

Catalog # Description MicroPulser Electroporator

165-2100 MicroPulser Electroporator, universal voltage, includes chamber with leads, 10 sterile cuvettes (5 packs of 0.1 cm and 0.2 cm gap)

**Cuvettes** 

165-2083 MicroPulser/Gene Pulser® Cuvettes, 0.1 cm gap, 5 (mini pack) 165-2089 MicroPulser/Gene Pulser Cuvettes, 0.1 cm gap, 50 (standard pack) 165-2093 MicroPulser/Gene Pulser Cuvettes, 0.1 cm gap, 500 (jumbo pack) 165-2082 MicroPulser/Gene Pulser Cuvettes, 0.2 cm gap, 5 (mini pack) MicroPulser/Gene Pulser Cuvettes, 0.2 cm gap, 50 (standard pack) 165-2086 165-2092 MicroPulser/Gene Pulser Cuvettes, 0.2 cm gap, 500 (jumbo pack) 165-2081 MicroPulser/Gene Pulser Cuvettes, 0.4 cm gap, 5 (mini pack) MicroPulser/Gene Pulser Cuvettes, 0.4 cm gap, 50 (standard pack) 165-2088 165-2092 MicroPulser/Gene Pulser Cuvettes, 0.4 cm gap, 500 (jumbo pack)

#### **Related Products and Information**



#### Cytofectene™ Transfection Reagent

Cytofectene is a powerful, ready-to-use cationic lipid transfection reagent. Cytofectene transfection reagent provides the highest transformation efficiencies with many cell types, high transformation efficiency in the presence of serum, minimal cytotoxicity, and a simple 1-step, 1-tube transformation procedure. Cytofectene is suitable for both adherent and suspension cultures and is effective for both transient and stable expression.



#### XenoWorks™ System

XenoWorks is a complete line of instrumentation designed for the rigorous demands of the latest microinjection and micromanipulation techniques. The system features ergonomic height-adjustable joystick controls, micromanipulator position memories, and variable movement radius. Microinjection, whether the delivery of DNA solution to a zygote's pronucleus, or insertion of embryonic stem cells into a blastocyst, can be achieved with a level of control previously unattainable with conventional instruments.



#### **Biolistics**

Biolistic technology, or particle bombardment, is a direct physical method of delivering nucleic acids or other molecules into cells. The Helios™ gene gun and PDS-1000/He™ system provide easy-to-use, rapid, versatile gene delivery that is independent of cell type and requires small amounts of DNA and few cells. This technology can be applied in vivo or in vitro to the widest range of targets, including cell cultures, tissues, organs, plants, and animals. These instruments use a helium pulse to accelerate high-density gold or tungsten particles coated with nucleic acids directly into target cells.



#### Electroporation

Electroporation is a highly efficient technique for introducing nucleic acids, proteins, and other molecules into a wide variety of cells. The Gene Pulser Xcell™ is a flexible, modular electroporator system that delivers exponential or square wave pulses optimal for your cell type. With manual and "optimize" capability, an intuitive interface, and pre-set programs, the Gene Pulser Xcell provides power and reliability.



**Bio-Rad** Laboratories, Inc.

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