

# Bio-Plex Pro™ TGF-β Assays

## Quick Guide

For Use with	Instruction Manual #
TGF-β Assays	10024984

This quick guide can be used to prepare and run a full 1 x 96-well assay plate. For more information on a given step, refer to the instruction manual. Go to [bio-rad.com/bio-plex](http://bio-rad.com/bio-plex) to download the instruction manual, which includes detailed instructions and a list of kit components.

**IMPORTANT!** Pay close attention to vortexing, shaking, and incubation instructions. Deviation from the protocol may result in low assay signal and assay variability.

## Initial Preparation

1. Plan the plate layout.
2. Start up/warm up the Bio-Plex® Suspension Array System (30 min).
  - Bring assay buffer, wash buffer, and sample diluent to room temperature (RT). Keep other items on ice until needed
  - Begin to thaw frozen samples
3. After thawing samples, activate them by adding 1 volume of 1 N HCl to 5 volumes of sample. Vortex, then incubate at RT for 10 min. Neutralize the samples by adding the same volume of base (1.2 N NaOH/0.5 M HEPES buffer). After treatment, dilute the samples according to the instructions in Table 1.

**Table 1. Dilution of samples.**

Sample Type	Diluent	Add Bovine Serum Albumin (BSA)	Sample Dilution
Serum and plasma	Sample diluent HB	None	1:16 final*
Culture media, with serum	Culture media	None	User optimized
Culture media, serum-free	Culture media	To 0.5% final	User optimized
Lavage, lysate, other fluids	Sample diluent HB	To 0.5% final	User optimized

\* For example, activate 25 µl sample, neutralize, and bring to a final volume of 400 µl.

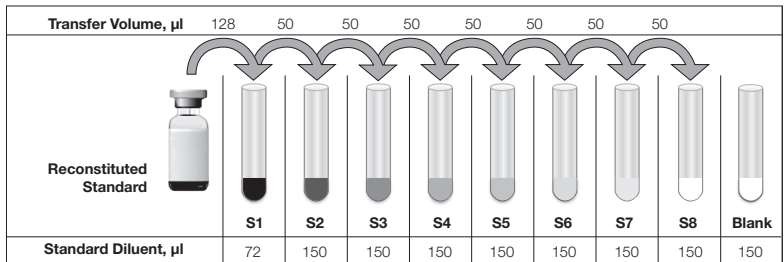
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- Prime the wash station for a flat-bottom plate. Prepare the 1x wash buffer. Mix the 10x stock by inversion to ensure all salts are in solution, then dilute 1 part 10x wash buffer (60 ml) with 9 parts distilled water (540 ml).
- Follow the prompts in Bio-Plex Manager™ Software to calibrate the system. This can be done now or during an incubation step.
- Mix 1 volume of standard diluent HB with 3 volumes of sample diluent HB (each supplied in the kit). The resulting solution is used for reconstitution and subsequent dilution of standards. This results in a serum matrix-based diluent that mimics the matrix in 1:16 diluted serum and plasma samples. For samples in serum-free media and other biological fluids, use a diluent that most closely matches the sample matrix. Add a carrier protein, such as BSA, at a final concentration of 0.5% (w/v).
- Reconstitute a single vial of standards in **500  $\mu$ l** of a diluent similar to the final sample type or matrix as shown in Table 2. **Vortex** for **5 sec** and **incubate on ice** for **30 min**.

**Table 2. Reconstitution of standards.**

Sample Type	Diluent for Standard	Add BSA
Serum and plasma	Standard/sample diluent HB mix (1:3)	None
Culture media, with serum	Culture media	None
Culture media, serum-free	Culture media	To 0.05% final (w/v)
Lavage, lysate, other fluids	Sample diluent HB	To 0.05% final (w/v)

- Prepare a fourfold standard dilution series and blank as shown in Figure 1. **Vortex** for **5 sec** between liquid transfers.



**Fig. 1. Schematic showing a fourfold dilution series and blank.**

- Vortex** the 20x coupled beads for **30 sec** and dilute to 1x in assay buffer (supplied in the kit) as shown in Table 3. Protect from light.

**Table 3. Dilution of coupled beads.**

Number of Wells	20x Beads, $\mu$ l	Assay Buffer, $\mu$ l	Total Volume, $\mu$ l
96	288	5,472	5,760

## Running the Assay

**Note:** Make sure all assay components are at RT before proceeding.

- Vortex** the diluted (1x) beads. Add **50  $\mu$ l** to each well of the assay plate.
- Wash two times with **100  $\mu$ l** wash buffer (supplied in the kit).
- Vortex** samples, standards, and blank. Add **50  $\mu$ l** to each well.
- Cover the plate with sealing tape. Incubate on a shaker at **850  $\pm$  50 rpm** for **2 hr** at RT.
- With **10 min** left in the incubation, **vortex** the 20x detection antibodies for **5 sec** and quick-spin to collect the liquid. Dilute to 1x in detection antibody diluent HB as shown in Table 4.

**Table 4. Dilution of detection antibodies.**

Number of Wells	20x Detection Antibody, $\mu$ l	Detection Antibody Diluent HB, $\mu$ l	Total Volume, $\mu$ l
96	150	2,850	3,000

- Wash the plate three times with **100  $\mu$ l** wash buffer.
- Vortex** the diluted (1x) detection antibodies. Add **25  $\mu$ l** to each well.
- Cover the plate with sealing tape and incubate on a shaker at **850  $\pm$  50 rpm** for **1 hr** at RT. Meanwhile, prepare the Bio-Plex Manager Software protocol; enter standard S1 values provided in the assay kit.
- With **10 min** left in the incubation, **vortex** the 100x streptavidin-phycoerythrin (SA-PE) for **5 sec** and quick-spin to collect the liquid. Dilute to 1x as shown in Table 5. Protect from light.

**Table 5. Dilution of SA-PE.**

Number of Wells	100x SA-PE, $\mu$ l	Assay Buffer, $\mu$ l	Total Volume, $\mu$ l
96	60	5,940	6,000

- Wash the plate three times with **100  $\mu$ l** wash buffer.
- Vortex** the diluted (1x) SA-PE. Add **50  $\mu$ l** to each well.

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- Cover the plate with sealing tape and incubate on a shaker at **850  $\pm$  50 rpm** for **30 min** at RT.
- Wash the plate three times with **100  $\mu$ l** wash buffer.
- Resuspend the beads in **125  $\mu$ l** assay buffer. Cover the plate with sealing tape and incubate on a shaker at **850  $\pm$  50 rpm** for **30 sec**.
- Remove the sealing tape and read the plate using the settings in Table 6.

**Table 6. Bio-Plex instrument settings.**

Instrument	RP1 (photomultiplier tube)	DD Gates	Bead Events
Bio-Plex 100, 200*	Low	5,000 (low), 25,000 (high)	50
Bio-Plex 3D*	Standard	Select MagPlex beads	50
Bio-Plex® MAGPIX™*	N/A, use default instrument settings		Default

\* Or similar Luminex system.

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