

Bio-Plex Pro Rat Diabetes Assays

Quick Guide

| For Use with | Instruction Manual # |
|---------------------|----------------------|
| Rat Diabetes Assays | 10000094509 |

This guide can be used to prepare and run a full 1 x 96-well assay plate. For more information on a given step, refer to the complete instruction manual. New users can go to bio-rad.com/bio-plex and download the manual, which includes detailed instructions and a list of kit components.

IMPORTANT! Pay close attention to **vortexing**, **shaking**, and **incubation** instructions. Deviation from the protocol may result in low assay signal and assay variability.










Initial Preparation

1. Plan the plate layout.
2. Start up/warm up the Bio-Plex System (**30 min**).
 - Bring assay buffer, wash buffer, and sample diluent to room temperature (RT). Keep other items on ice until needed
 - Begin to thaw frozen samples
 - Prepare 1x wash buffer. Mix 10x stock by inversion to ensure all salts are in solution. Then dilute 1 part 10x wash buffer (60 ml) with 9 parts distilled water (540 ml)
3. Prime the wash station for flat bottom plate.
4. Calibrate the Bio-Plex System by following the prompts in Bio-Plex Manager Software. This can be done now or during an assay incubation step.
5. Reconstitute a single vial of standards in **500 µl** of a diluent similar to the final sample type or matrix as shown below. **Vortex** for **5 sec** and incubate **on ice** for **30 min**.

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| Sample Type | Diluent for Standards | Add Bovine Serum Albumin (BSA) |
|---------------------------|-----------------------|--------------------------------|
| Serum and plasma | Standard diluent | None |
| Culture media, with serum | Culture media | None |
| Culture media, serum-free | Culture media | To 0.5% final (w/v) |

6. Prepare a fourfold standard dilution series and blank as shown below.
Vortex for 5 sec between liquid transfers.

| Transfer Volume, μ l | 128 | 50 | 50 | 50 | 50 | 50 | 50 | 50 | 50 | |
|---------------------------|---|---|---|---|---|---|---|---|---|-----|
| Reconstituted Standard |  |  |  |  |  |  |  |  |  | |
| Standard Diluent, μ l | 72 | 150 | 150 | 150 | 150 | 150 | 150 | 150 | 150 | 150 |

7. After thawing samples, prepare as shown below.

| Sample Type | Diluent | Add BSA | Sample Dilution |
|---------------------------|----------------|---------------------|-----------------|
| Serum and plasma | Sample diluent | None | Fourfold (1:4) |
| Culture media, with serum | Culture media | None | Neat to 1:10 |
| Culture media, serum-free | Culture media | To 0.5% final (w/v) | Neat to 1:10 |

Note: User-defined validation is required for the use of other dilution factors.

8. **Vortex** the 20x coupled beads for **30 sec** and dilute to 1x in Bio-Plex Assay Buffer as shown below. Protect from light.

| Number of Wells | 20x Beads, μ l | Assay Buffer, μ l | Total Volume, μ l |
|-----------------|--------------------|-----------------------|-----------------------|
| 96 | 288 | 5,472 | 5,760 |

Running the Assay

Note: Make sure all assay components are at RT before proceeding.

1. **Vortex** the diluted (1x) beads. Add **50 µl** to each well of the assay plate.
2. Wash the plate two times with **100 µl** Bio-Plex Wash Buffer.
3. **Vortex** samples, standards, and blank. Add **50 µl** to each well.
4. Cover the plate with sealing tape and protect from light with aluminum foil. Incubate on shaker at **850 ± 50 rpm** for **1 hr** at RT.
5. With 10 min left in the incubation, **vortex** the 20x detection antibodies for **5 sec** and quick-spin to collect liquid. Dilute to 1x in detection antibody diluent as shown below.

| Number of Wells | 20x Detection Ab, µl | Detection Ab Diluent HP, µl | Total Volume, µl |
|-----------------|----------------------|-----------------------------|------------------|
| 96 | 150 | 2,850 | 3,000 |

6. Wash the plate three times with **100 µl** wash buffer.
7. **Vortex** the diluted (1x) detection antibodies. Add **25 µl** to each well.
8. Cover the plate and incubate at **850 ± 50 rpm**, as in step 4, for **30 min** at RT. Meanwhile, prepare Bio-Plex Manager Software protocol; enter standard S1 values provided in the assay kit.
9. With 10 min left in the incubation, **vortex** the 100x streptavidin-phycoerythrin (SA-PE) for **5 sec** and quick-spin to collect liquid. Dilute to 1x as shown below and protect from light.

| Number of Wells | 100x SA-PE, µl | Assay Buffer, µl | Total Volume, µl |
|-----------------|----------------|------------------|------------------|
| 96 | 60 | 5,940 | 6,000 |

10. Wash the plate three times with **100 µl** wash buffer.
11. **Vortex** the diluted (1x) SA-PE. Add **50 µl** to each well.
12. Cover the plate and incubate at **850 ± 50 rpm**, as in step 4, for **10 min** at RT.

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13. Wash the plate three times with **100 µl** wash buffer.
14. Resuspend beads in **125 µl** assay buffer. Cover plate as in step 4 and shake at **850 ± 50 rpm** for **30 sec**.
15. Remove the sealing tape and **read the plate** using the settings below.

| Instrument | RP1 (PMT) | DD Gates | Bead Events |
|------------------|---|---|-------------|
| Bio-Plex 200* | High | 5,000 (low), 25,000 (high) | 50 |
| Bio-Plex 3D* | Enhanced | Select MagPlex Beads | 50 |
| Bio-Plex MAGPIX* | N/A, use default instrument settings | N/A, use default instrument settings | Default |

* Or similar Luminex System.

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