Quick Guide

**Standard Extraction Tube Protocol - Environmental Samples**

- Enrich the sample in buffered peptone water (for example 30 g in 270 ml), 18 hrs ± 2 hrs at 37°C
- Transfer 1 ml of enriched sample in a 1.5 ml screwcap tube

> Avoid including large fragments of food debris, and shaking stomacher bag before collecting

- Centrifuge at 10,000-12,000 g for 5 min
- Discard all the supernatant

- Add 200 μl of lysis reagent (reagent A)

> Lysis reagent must be constantly stirring in order to keep it in suspension
- Resuspend the pellet by pipetting the reagent up and down in the tube
- Vortex at high speed

- Incubate at 95-100°C for 10-15 min in a heating block
- Vortex at high speed
- Centrifuge at 10,000-12,000 g for 5 min

- Prepare the PCR mix
- Distribute 45 μl/well in the PCR microplate
- Add 5 μl of controls and sample supernatants

> Do not vortex before collecting the sample

> Check there are no bubbles
- Seal the microplate

- Start software
- Create the plate setup
- Start the amplification by clicking on “Run”

Please read the kit instruction manual and instrument user guide for complete and detailed instructions.
Enrich the sample in buffered peptone water (for example 30 g in 270 ml), 18 hrs ± 2 hrs at 37°C

Add 100 μl of lysis reagent (reagent A) in a 1.5 ml screwcap tube
Lysis reagent must be constantly stirring in order to keep it in suspension
Transfer 100 μl of enriched sample
Avoid including large fragments of food debris, and shaking stomacher bag before collecting

Mix by pipetting up and down and close the tube
Incubate at 95-100°C for 10-15 min in a heating block

Vortex at high speed
Centrifuge at 10,000-12,000 g for 2 min

Prepare the PCR mix
Distribute 45 μl/well in the PCR microplate
Add 5 μl of controls and sample supernatants
Do not vortex before collecting the sample
Check there are no bubbles
Seal the microplate

Start software
Create the plate setup
Start the amplification by clicking on “Run”