Quantifying fetal DNA in maternal blood plasma by ddPCR using DNA methylation

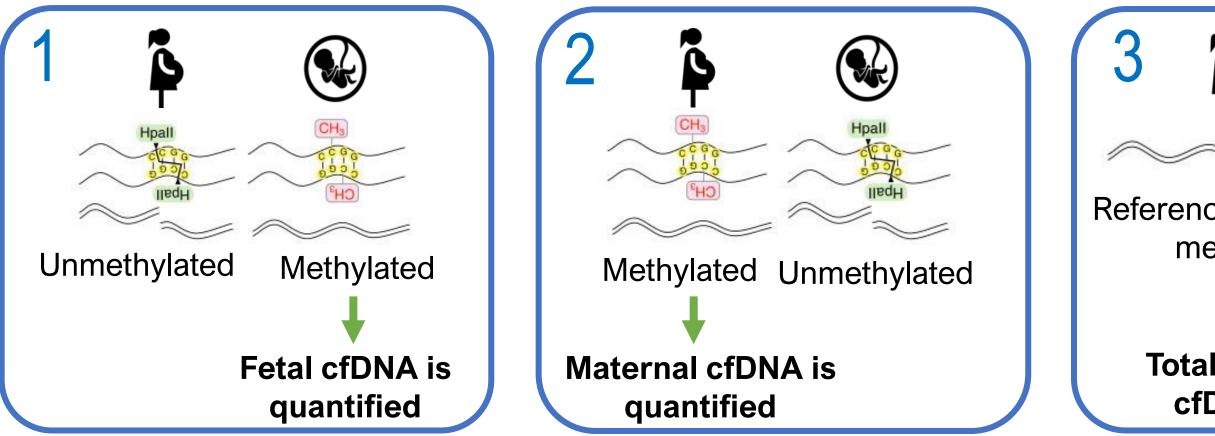
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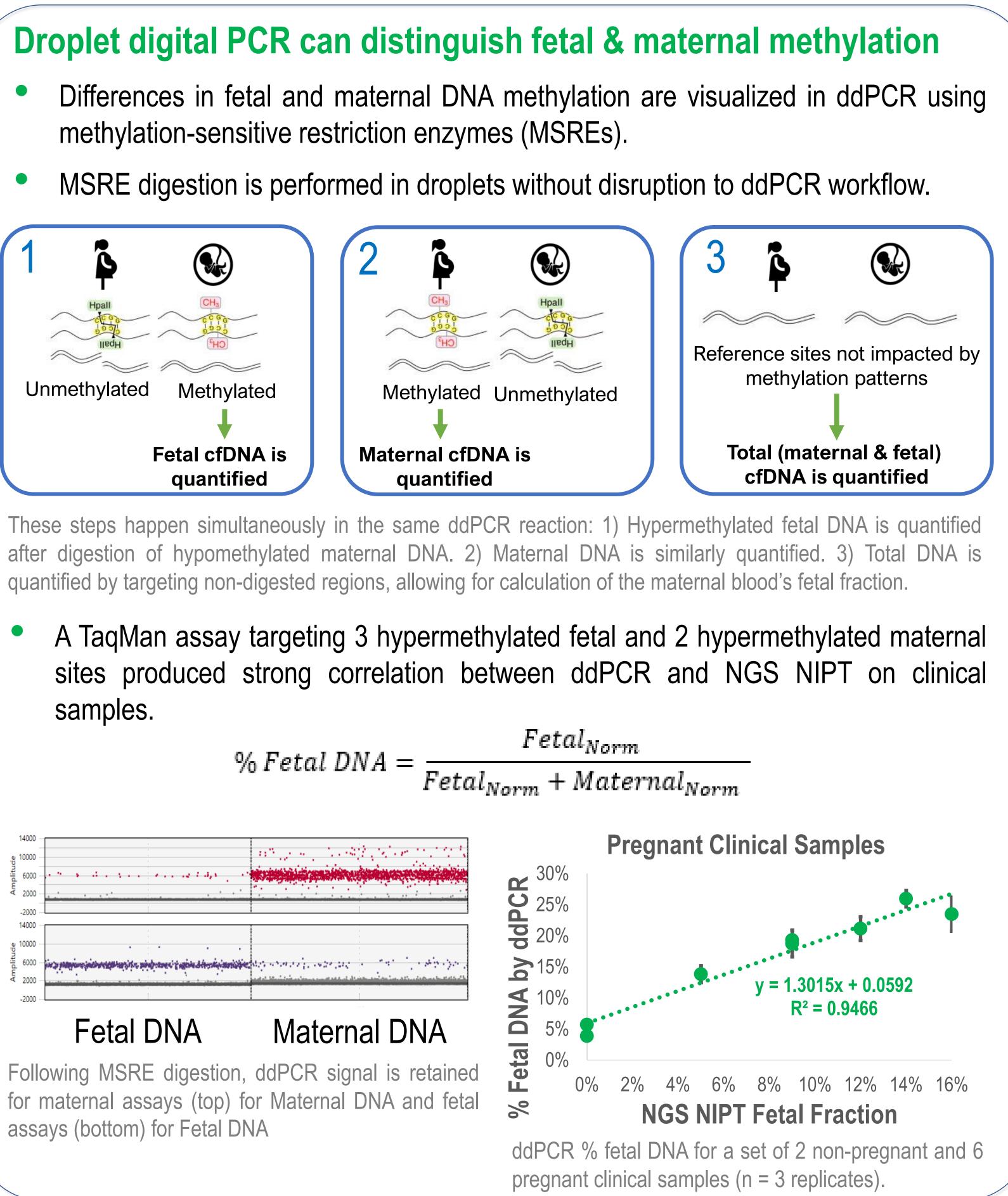
Abstract

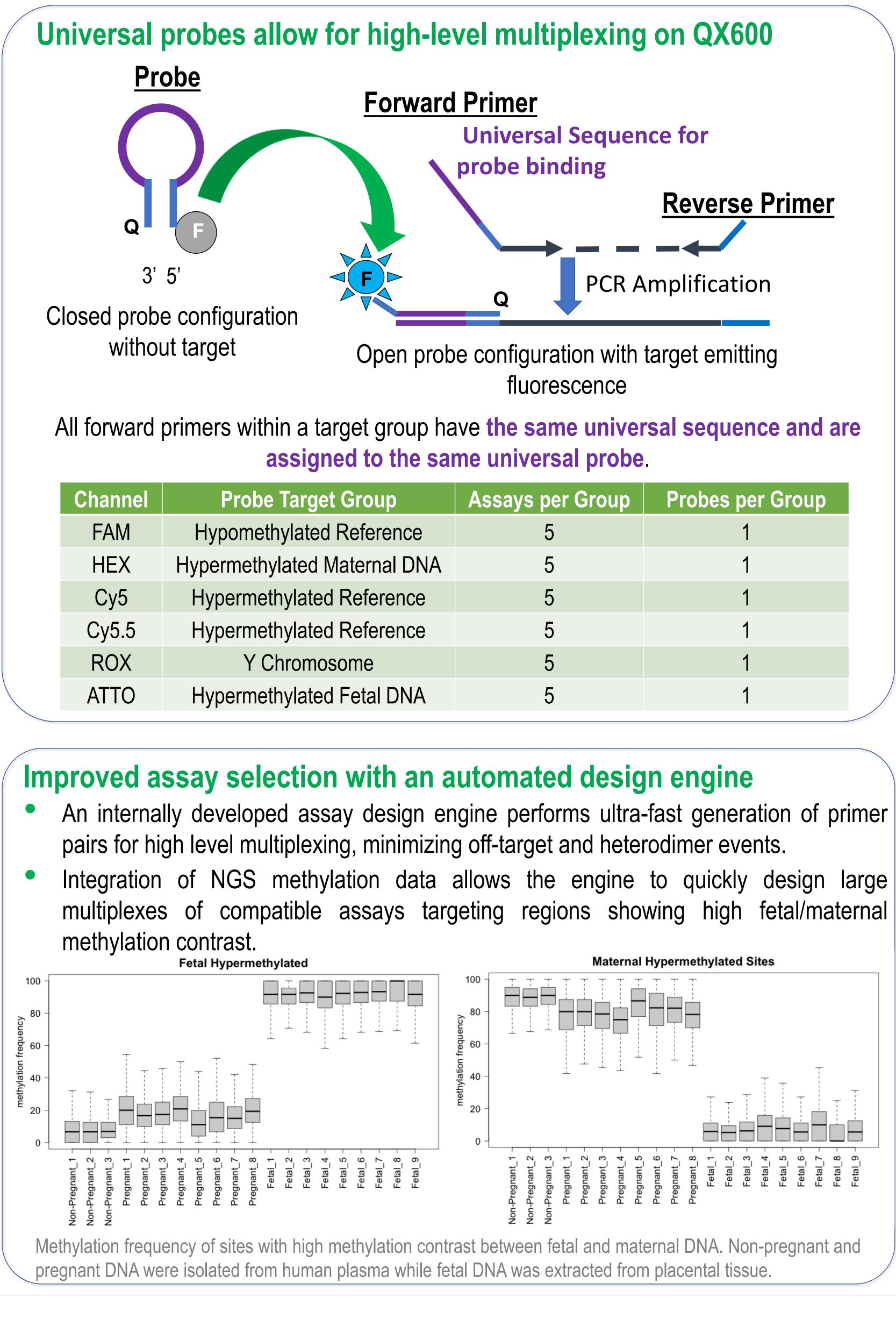
The proportion of cell-free DNA (cfDNA) circulating in maternal blood originating from the fetus, the fetal fraction, is an important quality control metric for tests on fetal-derived cfDNA. Epigenetic differences produce dissimilar DNA methylation patterns, and highcontrast regions are leveraged via methylation-sensitive digestion to quantify fetal and maternal DNA via droplet digital PCR (ddPCR). This advancement positions ddPCR as a faster and less expensive alternative to NGS for fetal fraction estimation.

- methylation-sensitive restriction enzymes (MSREs).



samples.

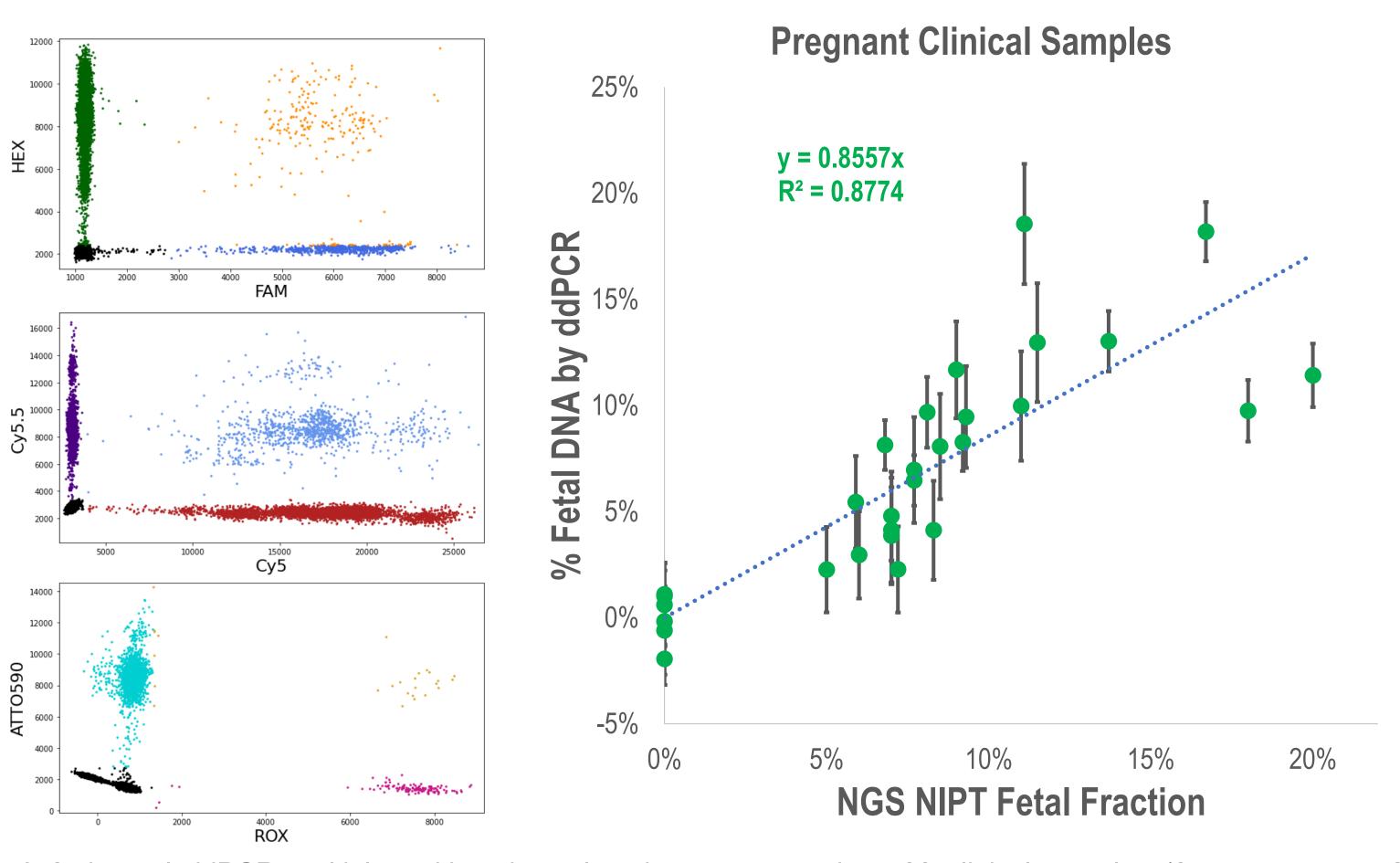




Assays per Group	Probes per Group
5	1
5	1
5	1
5	1
5	1
5	1

A ddPCR 30-plex for estimating fetal DNA in maternal blood plasma

- ddPCR.



A 6-channel ddPCR multiplex with universal probes was tested on 30 clinical samples (6 non-pregnant, 24 pregnant, n=1 replicate). 2D ddPCR plots are shown for one sample well with a 7% FF male pregnancy (left)). Samples showed good correlation with attached NGS NIPT results when corrected by non-pregnant samples.

Methylation-based analyses can be performed in ddPCR



A 30-plex (5 assays / target group) utilizing one universal probe per channel was tested on a set of 30 clinical samples on a Bio-Rad QX600 ddPCR system.

Positive droplet clusters were identified using an automated thresholding algorithm, after dynamic color calibration for each fluorescence channel.

Correlation (r² = 0.87) was observed between ddPCR fetal fraction and attached NGS NIPT values, following assay normalization and correction. Note: NGS NIPT FF values were obtained at earlier gestational ages than the sample collected for

Detection of methylation differences across otherwise similar DNA can be done with no impact to the ddPCR workflow.

• The use of universal probes allows for higher levels of multiplexing with only one probe used across several assays per fluorescence channel.

Integration of NGS methylation data with an automated high-plex design engine resulted in faster assay design targeting regions of high methylation contrast.

Efficient design of multiplexes targeting differentially methylated sites also has utility in other methylation-based biomarker analyses, including some cancers.

