



Genomic Variation Analysis: Single Nucleotide Polymorphism

Validation of Allele-Specific Expression Predicted by RNA-Seq in Human Brain Specimens

Yuka Imamura

Director, Penn State Hershey
Genome Sciences Facility
Assistant Professor of Pharmacology
Penn State College of Medicine*

“In our laboratory, we primarily use the ddPCR system to quantify absolute gene expression levels.”

Research Background

The goal of our research is to study genetic mechanisms contributing to the specialization of the human brain in the context of human development and disease occurrence. Technologies used in our laboratory include high-throughput sequencing, microarray, Droplet Digital PCR (ddPCR™), various bioinformatics tools, and gene manipulation in model organisms to functionally validate these findings. Allele-specific expression (ASE) is an important factor contributing to human phenotypic variability and the development of complex traits and diseases. We have employed RNA-Seq and SNP chip technology to predict ASE of 16 different brain regions from six healthy adult individuals.

Application

Among candidate alleles with an imbalanced expression from the six healthy individuals, one allele failed to be validated for its genotype by a standard quantitative PCR (qPCR) approach using an inventoried TaqMan assay. The homozygous A/A allele was not successfully resolved due to amplification of the G allele (likely occurring from nonspecific, cross-reactive amplification), generating ambiguous results (Figure 1). Several attempts with different settings yielded the same results. When I was going to move to some other technologies to overcome this problem, the QX100™ Droplet Digital PCR system was launched in our laboratory. We primarily use the ddPCR system to quantify absolute gene expression levels, especially for genes with low expression. I hoped the QX100 ddPCR system could also help assay genotype identification and allelic imbalance with higher precision.

ddPCR Results

As I expected, the QX100 ddPCR system generated a crystal clear, unambiguous result for genotype identification (Figure 2). Results show that the homozygous A/A allele generated nearly zero positive droplets using a G assay (FAM) while generating

twice as many droplets with an A assay (VIC) when the data were normalized by their input genomic content (Figure 2). I also saved our precious human genomic DNA (gDNA) by applying ten times less gDNA than the amount used for qPCR. The methods and instrument are robust, producing little variation in results among users once they are trained. I have applied ddPCR to quantify ASE events in cDNA samples obtained from various brain regions of different individuals and I achieved an absolute comparison of gene expression from both alleles (data not shown) without using any standard.

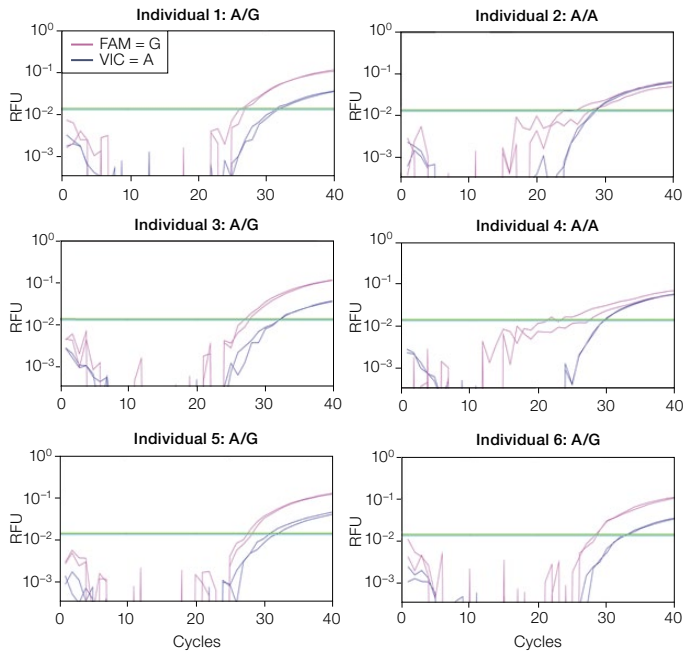


Fig. 1. Real-time PCR amplification curves for a candidate ASE allele. gDNA SNP assays were performed using qPCR. qPCR failed to call the A/A genotype. RFU, relative fluorescence units.

* Work was completed at Nenad Sestan Laboratory, Department of Neurobiology, Yale University School of Medicine.



Droplet Digital™ PCR Success Story

Sample	Description	Sample	Description
DO3	Individual 1 (A/G)	DO6	Individual 4 (A/A)
DO4	Individual 2 (A/A)	DO7	Individual 5 (A/G)
DO5	Individual 3 (A/G)	DO8	Individual 6 (A/G)

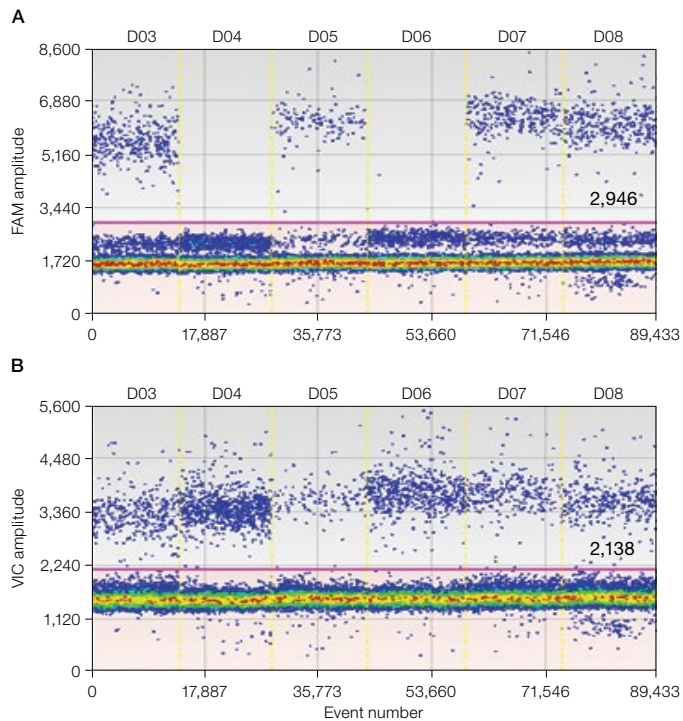


Fig. 2. Droplet Digital PCR validates both A/G and A/A genotypes. gDNA SNP assays were performed with G (A, FAM) and A (B, VIC) assays using ddPCR. A/G and A/A genotypes were successfully validated by ddPCR.

Conclusions

Bio-Rad's QX100 ddPCR system provides a method for accurately validating genotypes (SNPs). It also allows direct quantification of ASE (allelic imbalance) without the use of a standard, as opposed to standard qPCR that can provide indirect quantification. The costs and time constraints of ddPCR are negligible compared to other known methods, such as the SNaPshot multiplex system, for accurate ASE detection.

“Droplet Digital PCR outperforms qPCR in accurate genotype validation. It can also be used to quantify allele-specific expression events.”

Publication

Kang HJ et al. (2011). Spatio-temporal transcriptome of the human brain. *Nature* 478, 483–489.

FAM, SNaPshot, and VIC are trademarks of Applied Biosystems. TaqMan is a trademark of Roche Molecular Systems, Inc.

The QX100 Droplet Digital PCR system and/or its use is covered by claims of U.S. patents, and/or pending U.S. and non-U.S. patent applications owned by or under license to Bio-Rad Laboratories, Inc. Purchase of the product includes a limited, non-transferable right under such intellectual property for use of the product for internal research purposes only. No rights are granted for diagnostic uses. No rights are granted for use of the product for commercial applications of any kind, including but not limited to manufacturing, quality control, or commercial services, such as contract services or fee for services. Information concerning a license for such uses can be obtained from Bio-Rad Laboratories. It is the responsibility of the purchaser/end user to acquire any additional intellectual property rights that may be required.

For more information, visit

www.bio-rad.com/ddPCRSuccessMamura.



BIO-RAD

**Bio-Rad
Laboratories, Inc.**

Life Science
Group

Web site www.bio-rad.com USA 800 424 6723 Australia 61 2 9914 2800 Austria 01 877 89 01 Belgium 09 385 55 11 Brazil 55 11 5044 5699
Canada 905 364 3435 China 86 21 6169 8500 Czech Republic 420 241 430 532 Denmark 44 52 10 00 Finland 09 804 22 00
France 01 47 95 69 65 Germany 089 31 884 0 Greece 30 210 9532 220 Hong Kong 852 2789 3300 Hungary 36 1 459 6100 India 91 124 4029300
Israel 03 963 6050 Italy 39 02 216091 Japan 03 6361 7000 Korea 82 2 3473 4460 Mexico 52 555 488 7670 The Netherlands 0318 540666
New Zealand 64 9 415 2280 Norway 23 38 41 30 Poland 48 22 331 99 99 Portugal 351 21 472 7700 Russia 7 495 721 14 04
Singapore 65 6415 3188 South Africa 27 861 246 723 Spain 34 91 590 5200 Sweden 08 555 12700 Switzerland 026 674 55 05
Taiwan 886 2 2578 7189 Thailand 800 88 22 88 United Kingdom 020 8328 2000