If this is your first time using the Molecular Imager® system for protein stain imaging, we recommend purchasing Bio-Rad’s Oriole™ fluorescent gel stain (catalog # 161-0495) and using it with your samples on a standard Laemmli SDS-PAGE gel. Settings for Oriole gel stain are pre-programmed into the Image Lab software. Follow the steps shown in this quick start guide to ensure proper imaging of fluorescently stained protein gels.

**Application**
1. In the Gel Imaging step of the protocol set up, click on the Select button in the application section (Figure 1).
2. Select Protein Gels then choose the appropriate fluorescent stain from the drop-down menu (Figure 1).

**Imaging Area**
3a. Choose Select gel type, selecting from the drop-down menu if using Bio-Rad gels, in the Imaging Area dialog box (Figure 2).

OR

3b. Choose Enter image area and manually enter your gel dimensions in centimeters (W x L) (Figure 3).
**Image Exposure**

4a. Select the first option under **Image Exposure** if you would like the program to automatically optimize exposure time for **Intense** or **Faint** bands (Figure 4a).

OR

4b. If manual exposure time is desired, then choose **Manually set exposure time** and enter the desired exposure time (Figure 4b).

**Note:** Image Lab software will automatically display the exposure time of its last automated image next to the manual exposure time field as a guide for adjusting your image if you are not satisfied with the automated acquisition.

**Display Options**

5. Select your preference for the image display under **Display Options.** We recommend turning on **Highlight saturated pixels** and selecting **EtBr** from the **Image Color** drop-down menu (Figure 5).

Due to differential staining times and staining intensities, we recommend experimenting with several exposure times. To maximize the sensitivity and dynamic range of the imager, an optimal exposure time should be right below the point of saturation for the most concentrated protein bands.