

Instructions for Staining Polyacrylamide Gels

SDS-PAGE Mini Gels: 1) Wash gel 3 times for 5 min. each in 200 ml of ddH₂O per gel. 2) Remove all water from the staining container and add 50 ml Bio-Safe Coomassie Stain (or enough to completely cover gel). Gently shake for 1 hr. Protein bands will be visible within 20 min. and reach maximum intensity within 1 hr. Longer incubations will not increase background. 3) Rinse gel in 200 ml of ddH₂O for at least 30 min. Stained gels can be stored in water. **Note:** Residual SDS in the gel may cause background staining and interfere with band intensity while the gel is in the stain. Rinsing the gel extensively in water after staining will remove background and allow proper visualization of the bands.

SDS-PAGE Gels: For large gels (16-20") increase volumes in above method 2 fold.

PEPTIDE Gels: Fix gels in 40% methanol, 10% acetic acid for 30 mins., follow with staining starting with Step 2 (above). Extend Step 3 water wash to 2 hr. Peptide bands will not be clearly visible until after the final water wash.