

Gene Pulser® Electroprotocols

* We recommend adapting this protocol to use the Gene Pulser electroporation buffer (catalog #165-2676, 165-2677), which increases cell viability and transfection efficiency in mammalian cell lines.

Cell Type Mammalian, adherent Species Human, U373, glioblastoma, astrocytoma, grade III Used

Molecules Electroporated

DNA: multiple CAT and other vectors under different promoters including HIV, SIV, human and rhesus

cytomegalovirus immediate early gene, SV40 and the simian retrovirus type 2.

Before the Pulse

Cell growth medium RPMI 1640 with 10 mM dextrose

and 0.1mM dithiothreitol (Sigma, GIBCO/ BRL, Flow Labs)

Growth phase at harvest Pre-pulse

Log phase (We routinely subdivide cells 24 hours prior to electroporation

Trypsin harvest, washed twice in

Phosphate Buffered Saline

incubation

The Pulse

Wash solution

25°C Electroporation

Temperature

RPMI 1640 without Fetal Calf

Electroporation Medium* Serum, +10mM dextrose, 0.1 mM

dithiothreitol

1.3 x 10 (7) viable cells / ml, 0.3 ml **Cell Density**

 $300 \mu l$ Volume of Cells

 $500 \mu g / ml$ (5 to 10 μg per pulse) **DNA** Concentration

DNA Resuspension

Buffer

Not given

10 to 20 μ l Volume of DNA

Gene Pulser® apparatus & Instruments

None

Used Capacitance Extender

Cuvette Gap 0.4 cm

> Voltage 0.3 kV

Field 0.75 kV/cm Strength

960 μF Capacitor

Resistor (Pulse Controller) Ω none

Time 33 to 38 msec Constant

After the Pulse

DMEM culture media, 10% fetal calf **Outgrowth Medium**

serum

37 °C **Outgrowth Temperature** 48 hours Length of Incubation

Selection Method or

Assay Used

Electroporation

Transient (CAT, β -gal, immunohisto-chemistry).

50 to 100% Efficiency

Per Cent Survival 25 to 75 % Relevant Publications and/or Comments

Note: exponential values designated in parentheses. All plasmids were prepared by alkaline lysis and banded to equilibrium twice in cesium chloride gradients. Largest DNA molecule electroporated has been the rhesus cytomegalovirus genome (approx. 225 kilobases). In general, the higher the efficiency of transfection, the higher degree of cell killing. For some cell types,>90% of the input cells are killed, although > 90% of the survivors express very high levels of input DNA. We have found that $\geq 20 \mu g$ of DNA per electroporation resulted in greatly reduced efficiency of transfection. Similarly, 2.5 μ g of DNA resulted in reduced efficiency.

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