

Simultaneous Detection of 12 Phosphorylated and 7 Total Proteins Using the Bio-Plex Suspension Array System

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Introduction

Protein phosphorylation plays important roles in signal transduction, from activation of cell surface receptors to activation of transcription factors. To study intracellular signal transduction, it is important to detect the phosphorylation state of a target protein, which is directly related to its activity. To this end, the phosphorylation sites of many signal transduction proteins have been identified, and protein interactions have been profiled in a number of different pathways.

Phosphoprotein-specific antibodies are now available, enabling researchers to probe specific phosphorylated proteins. Traditionally, these types of investigations were carried out by western blot analysis, a lengthy process with relatively low throughput. More recently, the introduction of the Bio-Plex suspension array system, which uses an array of fluorescent microspheres and a dedicated reader with a digital processor (Figure 1), has allowed quantitation of up to 100 distinct analytes in a single sample.

We have developed a multiplex immunoassay panel that simultaneously measures the phosphorylation levels of 12 proteins (phospho-Akt, -ATF-2, -ERK, -GSK-3 α/β , -I κ B- α , -JNK, -p38 MAPK, -p70 S6 kinase, -p90RSK, -STAT2, -STAT3, and -TrkA) in a single well of a 96-well microplate filter. We also developed singleplex assays for phospho-EGFR and -S6. To gain a more comprehensive understanding of phosphorylation in a sample, we also developed a multiplex assay panel for simultaneous detection of 7 total target (phosphorylated and unphosphorylated) cell signaling proteins, Akt, ATF-2, ERK2, I κ B- α , JNK, p38 MAPK, and p90RSK. These Bio-Plex phosphoprotein and total target assays can be used, respectively, to measure the level of phosphorylation or total protein to generate up to 1,248 data points from a single 96-well plate.

Here we describe the performance of both types of multiplex assay panels, which have been validated with various cell lysates by correlation with western blots.

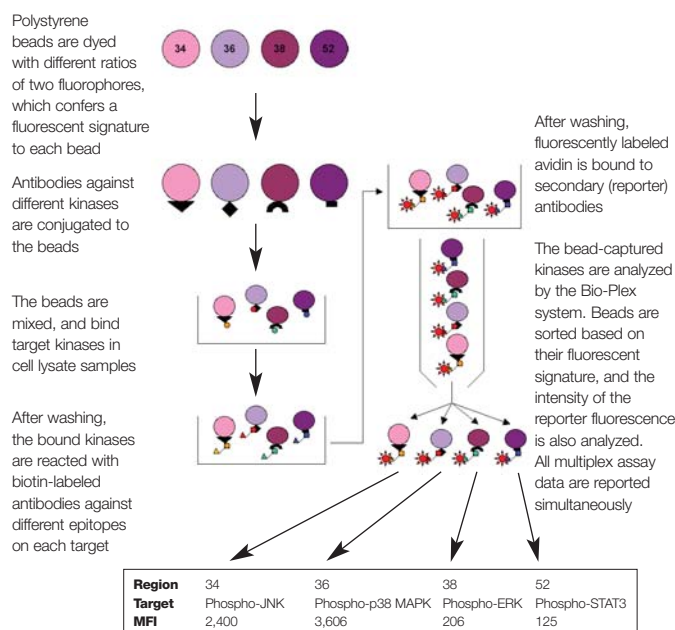


Fig. 1. Bio-Plex phosphoprotein assays.

Table 1. Target information.

Target Protein	Antibody-Reactive Species	Cell Lysate Species Tested	Phosphorylation Site(s)
Phospho-Akt	Human, mouse, rat	Human, mouse	Ser ⁴⁷³
Phospho-ATF-2	Human, mouse, rat	Human, mouse	Thr ⁷¹
Phospho-EGFR*	Human	Human, mouse	Tyr
Phospho-ERK1/2	Human, mouse, rat	Human, mouse	Thr ²⁰² /Tyr ²⁰⁴ , Thr ¹⁸⁵ /Tyr ¹⁸⁷
Phospho-GSK-3 α/β	Human, mouse, rat	Human, mouse	Ser ²¹ /Ser ⁹
Phospho-I κ B- α	Human, mouse	Human, mouse	Ser ³² /Ser ³⁶
Phospho-JNK	Human, mouse	Human, mouse	Thr ¹⁸³ /Tyr ¹⁸⁵
Phospho-p38 MAPK	Human, mouse, rat	Human, mouse	Thr ¹⁸⁰ /Tyr ¹⁸²
Phospho-p70 S6 kinase	Human, mouse, rat	Human, mouse	Thr ⁴²¹ /Ser ⁴²⁴
Phospho-p90RSK	Human, mouse, rat	Human, mouse	Thr ²⁵⁹ /Ser ³⁸³
Phospho-S6*	Human, mouse, rat	Human, mouse	Ser ²³⁵ /Ser ²³⁶
Phospho-STAT2	Human	Human	Tyr ⁶⁸⁹
Phospho-STAT3	Human, mouse, rat	Human, mouse	Tyr ⁷⁰⁵
Phospho-TrkA	Human, mouse, rat	Human, rat	Tyr ⁴⁹⁶

* Bio-Plex singleplex assay

Methods

Treated and untreated control cultures were prepared at the same time. Cultured cells were lysed according to the instructions specified in the Bio-Plex cell lysis kit product insert. An equal volume of assay buffer from the Bio-Plex phosphoprotein reagent kit was added to each sample. The cell culture lysates were analyzed according to the instructions specified in the Bio-Plex phosphoprotein detection instruction manual. The protocols for lysate preparation and assays are summarized in Figure 2.

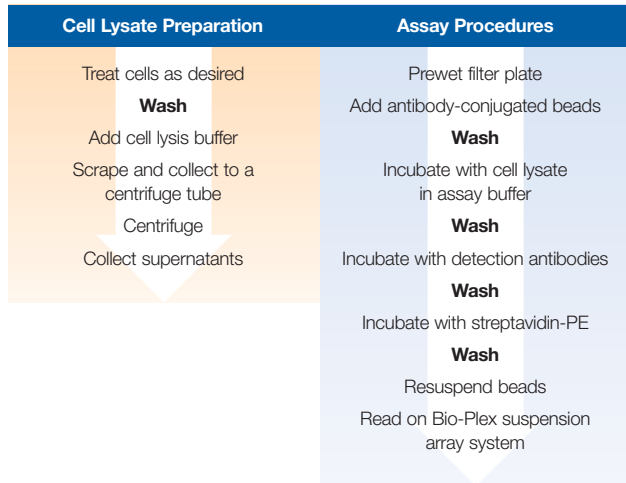


Fig. 2. Protocols for lysate preparation and assays.

Results and Discussion

Table 1 identifies the target proteins investigated here and their respective phosphorylation sites. Bio-Plex assays were tested for correlation with western blots for 14 phosphoprotein targets using lysates from different treated cell lines. The cell lysates were diluted, and for each dilution, the same amount of total protein (μg per well or lane) was tested both by Bio-Plex assay and western blotting. Figure 3 summarizes the results. A good dose response was demonstrated for each of the 14 diluted targets, and the mean fluorescence intensity (MFI) from the Bio-Plex phosphoprotein assays correlated very well with the band intensities seen in western blots.

Bio-Plex total target assays were also compared to western blots for serially diluted untreated HEK 293, HeLa, or NIH 3T3 cell lysates. For each dilution, the same amount of total protein (μg per well or lane) was tested both by Bio-Plex assay and western blotting. Figure 4 shows the correlation between the Bio-Plex total target assay MFI values and intensity of the bands on the western blots. All 7 of the Bio-Plex total target assays demonstrated a linear dose-response and good correlation with western blots.

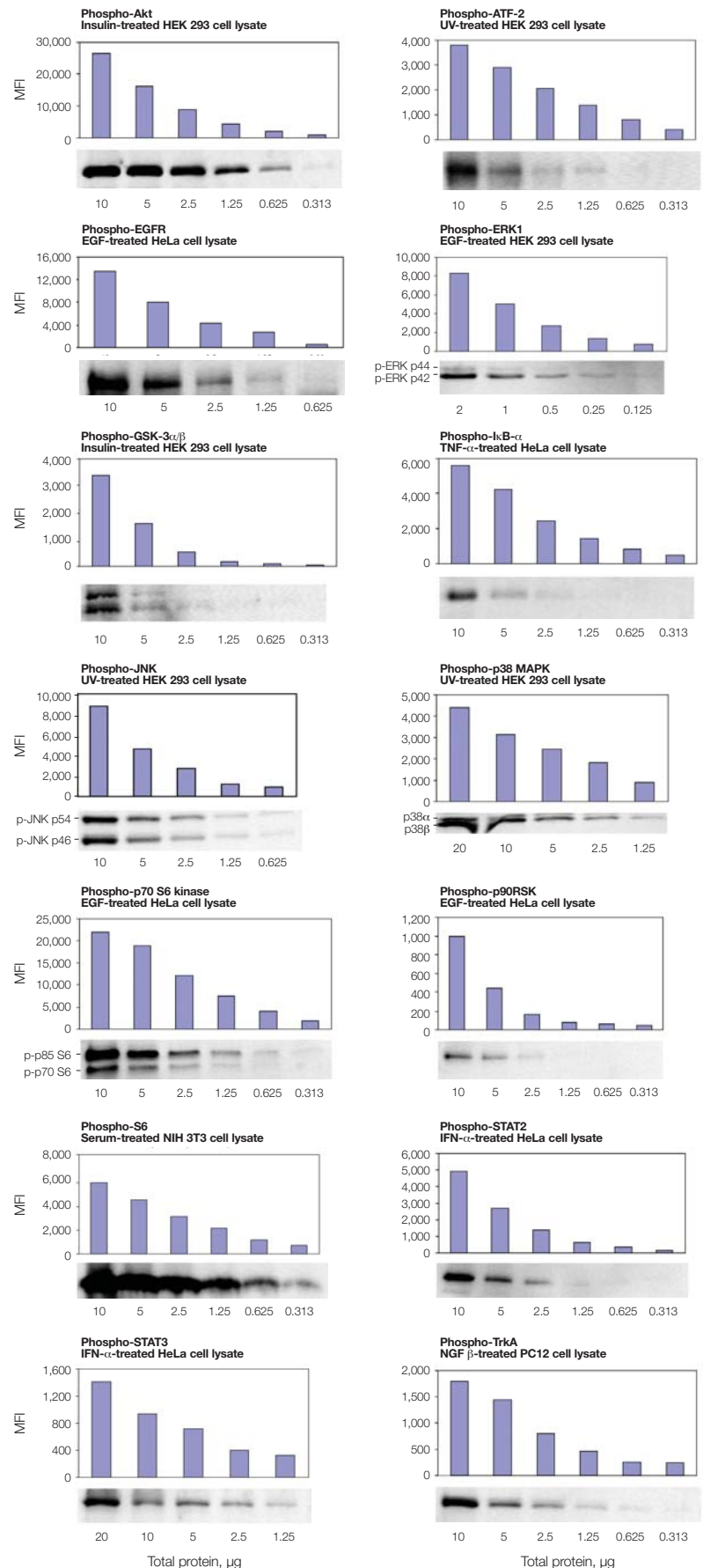


Fig. 3. Correlation of Bio-Plex phosphoprotein assays (upper panels) with western blots (lower panels). Note correlation of assay MFI values to band intensities. To compensate for the low-level phosphorylation of phospho-ATF-2 in cultured cells, mouse tissue expressing a high level of ATF-2 was used.

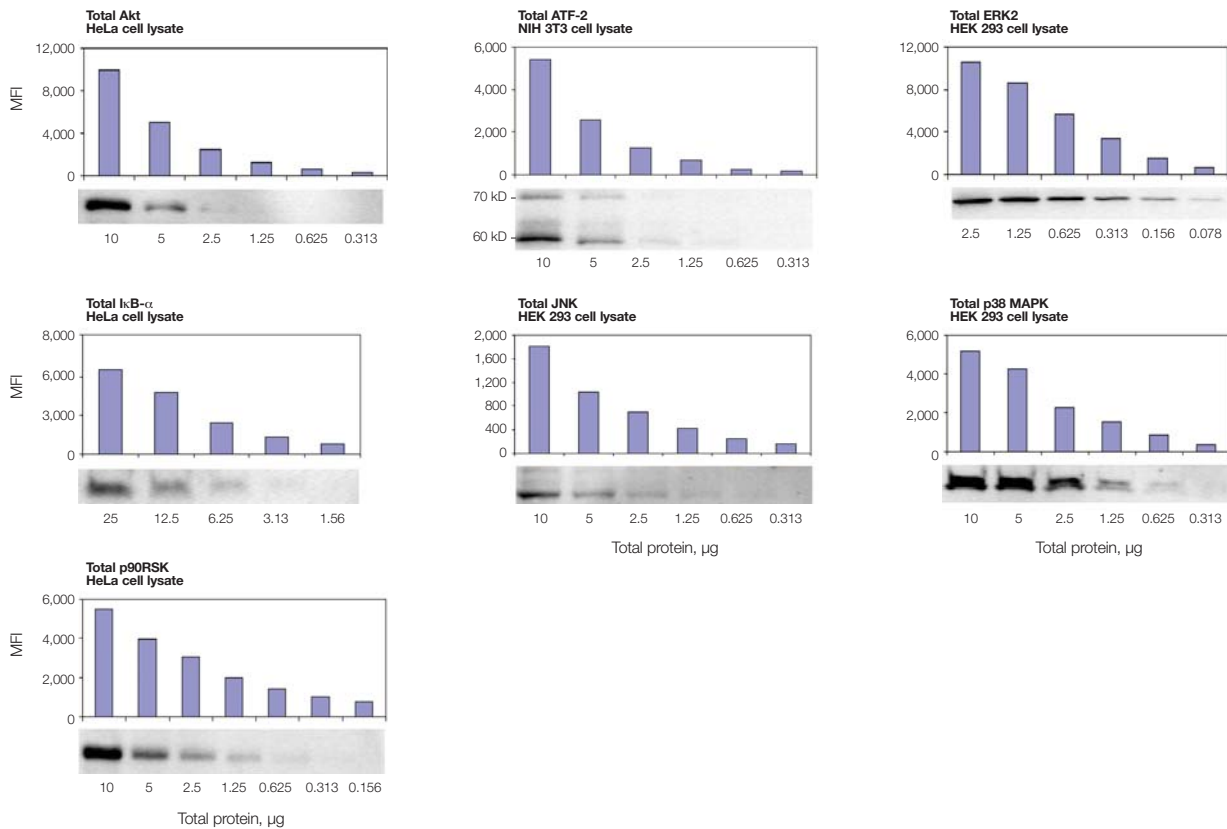


Fig. 4. Correlation of Bio-Plex total target assays (upper panels) with western blots (lower panels). Note correlation of assay MFI value to band intensities.

Conclusions

- The multiplex Bio-Plex phosphoprotein or total assays detect different phosphorylated or total targets simultaneously from one well of a 96-well plate
- Currently, Bio-Plex assays are available for 12 multiplexable phosphoprotein and 7 multiplexable total assays
- Bio-Plex singleplex assays for phospho-EGFR and phospho-S6 have also been developed
- These multiplex phosphoprotein assay panels exhibit greater or equal sensitivity compared to western blots

The Bio-Plex suspension array system includes fluorescently labeled microspheres and instrumentation licensed to Bio-Rad Laboratories, Inc. by the Luminex Corporation.

Information in this tech note was current as of the date of writing (2004) and not necessarily the date this version (Rev B, 2005) was published.



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