

# Aurum™ Plasmid Mini Kit: Spin Format



## Protocol Overview

For complete protocol, consult instruction manual.

### Growth and Isolation

1. Grow 1–5 ml bacterial culture overnight or  $\geq 16$  hr.
2. Measure  $A_{600}$  (if higher yield required).
3. Transfer an appropriate volume of culture to a capped 2 ml tube. Centrifuge and decant supernatant.
4. Add 250  $\mu$ l resuspension solution; vortex.
5. Add 250  $\mu$ l lysis solution; invert 6–8x.
6. Add 350  $\mu$ l neutralization solution; invert 6–8x.
7. Centrifuge 5 min to pellet cell debris.

250  $\mu$ l  
resuspension  
solution



Vortex

250  $\mu$ l  
lysis solution



Invert 6–8x

350  $\mu$ l  
neutralization  
solution



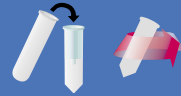
Invert 6–8x



### Purification

8. Transfer cleared lysate (supernatant) to mini spin/vac column.
9. Centrifuge 1 min to bind plasmid DNA. Decant flow-through.
10. Add 750  $\mu$ l wash solution and centrifuge 1 min. Decant flow-through.
11. Centrifuge additional **1 min** to remove residual wash solution.

Transfer cleared lysate



750  $\mu$ l  
wash  
solution



### Collection of Purified Samples

12. Transfer mini spin/vac column to a clean 1.5–2.0 ml capped tube.
13. Add 50  $\mu$ l elution solution. Let stand 1 min and then centrifuge 1 min to elute.
14. Purified DNA is ready to use or can be stored at 4°C.

50  $\mu$ l  
elution  
solution



**BIO-RAD**

Aurum Plasmid Mini Kit: Cat. #732-6400

For more information, call Technical Services at 1-800-4BIORAD (1-800-424-6723). Visit us on the Web at [www.bio-rad.com/aurum/](http://www.bio-rad.com/aurum/)

# Aurum™ Plasmid Mini Kit: Vacuum Format



## Protocol Overview

For complete protocol, consult instruction manual.

### Growth and Isolation

1. Grow 1–5 ml bacterial culture overnight or  $\geq 16$  hr.
2. Measure  $A_{600}$  (if higher yield required).
3. Transfer an appropriate volume of culture to a capped 2 ml tube. Centrifuge and decant supernatant.
4. Add 250  $\mu$ l resuspension solution; vortex.
5. Add 250  $\mu$ l lysis solution; invert 6–8x.
6. Add 350  $\mu$ l neutralization solution; invert 6–8x.
7. Centrifuge 5 min to pellet cell debris.

250  $\mu$ l  
resuspension  
solution



Vortex

250  $\mu$ l  
lysis solution



Invert 6–8x

350  $\mu$ l  
neutralization  
solution



Invert 6–8x



Transfer  
cleared lysate

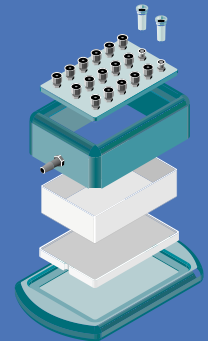


### Purification on Aurum or Comparable Manifold

(See exploded view for proper setup of manifold.)

8. Transfer cleared lysate (supernatant) to mini spin/vac column.
9. Apply vacuum at -20 to -23" Hg to bind plasmid DNA. Turn vacuum off.
10. Add 750  $\mu$ l wash solution and reapply vacuum until all liquid has passed through column.

750  $\mu$ l  
wash solution



11. Transfer mini spin/vac column to a 2 ml wash tube. Spin **1 min** to remove residual wash.



### Collection of Purified Samples

12. Transfer mini spin/vac column to a clean 1.5–2.0 ml capped tube.
13. Add 50  $\mu$ l elution solution. Let stand 1 min and then centrifuge 1 min to elute.
14. Purified DNA is ready to use or can be stored at 4°C.

50  $\mu$ l  
elution  
solution



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