ELECTROPHORESIS

Instructions for Staining Polyacrylamide Gels

Quick Start Guide

SDS-PAGE Mini Gels

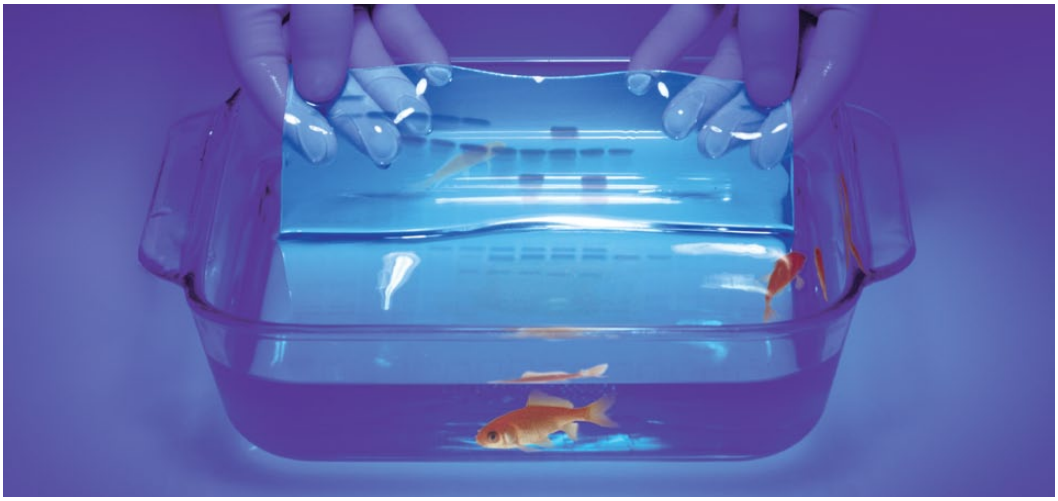
1. Wash the gel 3 times for 5 min each in 200 ml of ddH_2O per gel to remove SDS, which will interfere with the staining.
2. Remove all the water from the staining container and add 50 ml Bio-Safe™ Coomassie stain (or enough to completely cover the gel).
3. Gently shake for 1 hr. Protein bands will be visible within 20 min and reach maximum intensity within 1 hr. Longer incubations will not increase background.
4. Rinse the gel in 200 ml of ddH_2O for at least 30 min. Rinsing the gel extensively in water after staining will further reduce background. Stained gels can be stored in water.

SDS-PAGE Large Format Gels

For large gels (16–20") increase volumes in the above protocol twofold.

Peptide Gels

Fix gels in 40% methanol, 10% acetic acid for 30 min and follow the above protocol starting with step 2. Extend step 3 water wash to 2 hr. Peptide bands will not be clearly visible until after the final water wash.



Ordering Information

Catalog #	Description
161-0786	**Bio-Safe Coomassie Stain**, 1 L
161-0787	**Bio-Safe Coomassie Stain**, 5 L

BIO-RAD