

# **Bio-Plex Pro Mouse Th17 Cytokine Assays**

#### **Quick Guide**

For Use with	Instruction Manual #
Bio-Plex Pro Cytokine, Chemokine, and Growth Factor Assays	10000111560

This guide can be used to prepare and run a full 1 x 96-well assay plate. New users should download the complete manual, which includes detailed instructions and a list of kit components, at bio-rad.com/bio-plex.

#### **Initial Preparation**

- 1. Plan the plate layout.
- 2. Start up/warm up the Bio-Plex Multiplex Immunoassay System (30 min).
  - Bring diluents, including wash buffer, assay buffer, standard diluent, detection antibody diluent HP, and sample diluent, to room temperature (RT). Keep the other items on ice until needed
    - Mix by inversion to ensure all salts are in solution
    - Prepare 1x wash buffer: dilute 1 part 10x wash buffer (60 ml) with 9 parts distilled water (540 ml)
  - Begin to thaw the frozen samples
- 3. Prepare the sample dilution according to the guidelines provided in the following table. It is important to centrifuge serum or plasma samples at 1,000 x g for 15 min at 4°C to remove particulates from all samples prior to use.

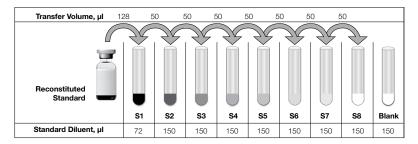
	Serum and Plasma		Culture Supernatant and Other Fluids		Cell and Tissue Lysate	
Assay	Dilution	Diluent	Dilution	Diluent	Dilution	Diluent
Mouse and rat cytokines	1:4	Bio-Plex Sample Diluent	User optimized	Cell culture medium or buffer similar to sample*	User optimized (1:2 of lysates at 200–900 µg/ml protein)	Bio-Plex Sample Diluent
Mouse ICAM-1	1:100	Bio-Plex Standard and Sample Diluents	User optimized			

<sup>\*</sup> If samples are serum-free, add bovine serum albumin (BSA) to 0.5% final w/v.

## For example:

- For serum or plasma cytokine assays, dilute samples 1:4 by adding 40 μl sample + 120 μl Bio-Plex Sample Diluent
- For serum or plasma ICAM-1 assays, dilute samples 1:100
  - First dilution (1:4): 10 μl sample + 30 μl Bio-Plex Sample Diluent
  - Second dilution (1:25): 5 μl from the first dilution + 120 μl Bio-Plex Standard Diluent
- 4. Calibrate the Bio-Plex System in Bio-Plex Manager Software.
- 5. Reconstitute the standards and control by adding 500 μI of standard diluent to each. Vortex at medium speed for 5 sec and incubate all vials on ice for precisely 30 min.
- 6. Prepare a fourfold standard dilution series and blank as shown. **Vortex** at medium speed for **5 sec** between liquid transfers.

Note: The control is ready to use after reconstitution. Controls are included with the fixed panel only.



 Vortex the coupled beads at medium speed for 30 sec and dilute to 1x in Bio-Plex Assay Buffer as shown. Protect from light.

#### **Premixed Panels**

Number of Wells	10x Beads, μΙ	Assay Buffer, μl	Total Volume, μΙ
96	570	5,130	5,700

#### Singleplex Assays

Number of Wells	Singleplex #1 10x Beads, μΙ	Singleplex #2 10x Beads, μΙ	Assay Buffer, µl	Total Volume, μΙ
96	570	570	4,560	5,700

Note: 10x singleplex beads allow multiplexing up to ten analytes.

#### **Running the Assay**

- 1. Vortex the diluted (1x) beads. Add 50 µl to each well of the assay plate.
- 2. Wash the plate two times with 100 µl Bio-Plex Wash Buffer.
- 3. Vortex the samples, standards, blank, and control. Add 50  $\mu l$  to each well.
- 4. Cover the plate with sealing tape. Incubate on shaker at  $850 \pm 50$  rpm at RT for 30 min.
- 5. With 10 min left in the incubation, **vortex** the detection antibodies for **5 sec** and quick-spin to collect liquid. **Dilute to 1x** as shown.

#### **Premixed Panels**

Number of Wells	10x Detection Antibodies, µl	Detection Antibody Diluent HP, μl	Total Volume, µl
96	300	2,700	3,000

## Singleplex Assays

Number of	10	x Detection 10		tection Antibody Diluent HP, µl T	otal Volume, µl
96		300	300	2,400	3,000

Note: 10x singleplex beads allow multiplexing up to ten analytes.

- **6. Wash the plate three times** with **100 μl** wash buffer.
- 7. Vortex the diluted (1x) detection antibodies. Add 25  $\mu l$  to each well.
- 8. Cover the plate with sealing tape and incubate at  $850 \pm 50$  rpm for 30 min at RT. Meanwhile, prepare the Bio-Plex Manager Software protocol; enter standard S1 values and units provided in the assay kit.
- **9.** With 10 min left in the incubation, **vortex** 100x streptavidin-phycoerythrin (SA-PE) for **5 sec** and quick-spin to collect liquid. **Dilute to 1x** as shown and protect from light.

Number of Wells	100x SA-PE, μI	Assay Buffer, µl	Total Volume, µl
96	60	5,940	6,000

- 10. Wash the plate three times with 100 µI wash buffer.
- 11. Vortex the diluted (1x) SA-PE. Add 50 µI to each well.
- 12. Cover the plate with sealing tape and incubate at  $850 \pm 50$  rpm for 10 min at RT.
- 13. Wash the plate three times with 100  $\mu I$  wash buffer.
- 14. Resuspend the beads in 125  $\mu$ I assay buffer. Cover and shake at 850  $\pm$  50 rpm for 30 sec.
- **15.** Remove the sealing tape and **read plate** using the following settings:

Instrument	RP1 (PMT)	DD Gates	Bead Events
Bio-Plex 3D*	Standard	Select MagPlex Beads	50
Bio-Plex 100, 200*	Low	5,000 (low); 25,000 (high)	50
Luminex MAGPIX	N/A, use default instrument settings		

<sup>\*</sup> Or similar Luminex System.

### **Assay Workflow**

Add 50 μl 1x beads to wells
<u></u>
Wash buffer: 2 x 200 µl
<u> </u>
Add 50 µl standards, samples, controls; incubate on shaker at 850 rpm for 30 min at RT
<b>→</b>
Wash buffer: 3 x 100 μl
<b>★</b>
Add 25 µl 1x detection antibody; incubate on shaker at 850 rpm for 30 min at RT
<u> </u>
Wash buffer: 3 x 100 µl
<b>→</b>
Add 50 µl 1x SA-PE; incubate on shaker at 850 rpm for 10 min at RT
<u> </u>
Wash buffer: 3 x 100 μl
<b>→</b>
Resuspend in 125 µl assay buffer; shake at 850 rpm for 30 sec
<b>→</b>
Acquire data on Bio-Plex System
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