

Affi-Gel® Boronate Affinity Gel to Separate Ribonucleotides, Ribonucleosides, Sugars, Catecholamines, and Coenzymes

Introduction

Low molecular weight molecules such as nucleotides, nucleosides, catecholamines, coenzymes, and sugars can be separated very efficiently by chromatography on Affi-Gel boronate gel. Affi-Gel boronate gel is a boronate-derivatized polyacrylamide gel with an affinity for adjacent *cis* hydroxyl (*cis*-diol) groups (Figure 1). It effectively separates molecules, such as AMP and cyclic AMP (cAMP), which differ only in the presence or absence of such a group. It also separates *cis*-diol-containing molecules such as cytosine, uridine, and adenosine from one another — all are bound, but their differing affinities permit separate elution (Figure 2A). Bio-Gel® P-6 beads, the gel beads derivatized to make Affi-Gel boronate gel, have a molecular weight exclusion limit of 6,000. The binding capacity of Affi-Gel boronate gel is, therefore, extremely high for low molecular weight compounds.

The development and use of boronate affinity media stems from the observation that boronate forms complexes with sugars possessing coplanar *cis*-diol groups (Boeseken 1949). The structure of the boronate affinity group in Affi-Gel boronate gel is:

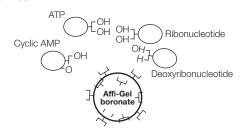
The mechanism of binding has been suggested to be reversible formation of a five-member ring (Lorand et al. 1959).

Affi-Gel boronate

Free diol Bound dio

Boronate derivatives on various solid supports have been investigated and applied (Table 1). These derivatives have been variously termed dihydroxy boryl, boric acid gel, and boronic acid gel, all referring to m-aminophenyl boronic acid immobilized to solid supports. Supports have included aminoethyl cellulose, porous glass, methacrylate gel, and Bio-Gel P polyacrylamide gel.

Sample application



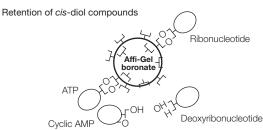


Fig. 1. Separation by affinity for cis-diols.

Affi-Gel boronate gel offers a variety of advantages as a support for the applications listed in Table 1:

- High boronate capacity 1.05 ± 0.15 meq/g
- High capacity for diols binds 130 µmol sorbitol/ml gel*
- Spherical polyacrylamide matrix excellent flow rate, resolution, and reproducibility
- Low nonspecific absorption, optimum porosity low exclusion limit precludes significant binding of high molecular weight molecules, while allowing rapid equilibration and binding of low and medium molecular weight molecules

Preparation and Use

The gel is supplied as dried material and must be hydrated in an appropriate buffer. Buffer choice is dependent upon application (Table 1). Buffers such as Tris should be avoided, since they can adversely affect binding capacity (Schott et al. 1973). The presence of Mg²⁺ may enhance binding. In general, binding is effected at pH >7.5 and elution at pH <6.5. Boric acid, sorbitol, or mannitol can also be used for elution.

* Dr R Hurlbert, personal communication.



Table 1. Applications using boronate affinity media.

| Application | Application Buffer | Molecules in ${\rm V_o}$ | Retained Molecules | Elution Buffer | Support Used | References |
|---|--|--|--|--|---|--|
| Adenylate cyclase assay | HEPES pH 7.5 + MgCl ₂ | cAMP | ATP, AMP, and adenosine | 0.05 M NaOAc | Bio-Gel P-150 boronate gel | Hageman and Kuehn 1977 |
| Isolation of catecholamines from urine | 0.1 M phosphate pH 7.0 + EDTA | Other urine components and DOPA | Norepinephrine, epinephrine | 0.025 N HCI | Boric acid gel (Sigma-Aldrich) | Higa et al. 1977 |
| Modified nucleosides in urine | 0.25 M NH ₄ OAc, pH 8.8 | Thymidine, adenine | Pseudouridine | 0.1 M HCOOH | Bio-Gel P-2 gel, 200–400 mesh, boronate | Davis et al. 1977 Uziel et al. 1976 |
| Separation of mono- and oligonucleotides | Triethyl ammonium | Deoxyribo- nucleotides | Ribonucleotides | H ₂ O and others | Dihydroxyboryl methacrylate | Schott et al. 1973 |
| Sugars | 0.05 M N-methyl- morpholinium-CI, pH 7.5 + 1 M NaOAc | Erythritol, adonitol, sucrose, and D-glucose | L-arabinitol, xylitol, D-mannitol, dulcitol, sorbitol, and D-fructose* | Isocratic run, elution buffer same as application buffer | Dihydroxyboryl cellulose | Weith et al., 1970 |
| Assay of ribonucleotide reductase | Tris, MgCl ₂ | dUMP, dCMP | CDP | Citrate | Dihydroxyboryl cellulose Dowex 1(AG) resin | Moore et al. 1974 |

^{*} In order of increasing retention by support.

Regeneration and Storage

Regeneration of the gel is facilitated by washing with starting buffer if a low-pH elution buffer has been used. If boric acid buffer or diol solutions have been used for elution, the gel must first be washed with 0.1 M acetic acid before reequilibrating with starting buffer.

Hydrated gel can be stored at 4°C under neutral pH conditions for up to 2 months; dry material has excellent long-term stability.

Typical Procedure

Stepwise elution is typical for separation of a ribonucleotide (such as GMP) from a cyclic nucleotide (cGMP) (Figure 2B):

- 1. Prepare a 1 x 4 cm column of Affi-Gel boronate in 0.1 M HEPES, pH 8.5.
- 2. Load a 200 μl solution containing 1 μmol each cGMP and GMP in the starting buffer onto the column.
- Wash the column with 5 column volumes of starting buffer at a flow rate of 1 ml/min and collect the cyclic nucleotide.
- 4. Elute the ribonucleotide with 0.1 M NaPO₄, pH 6.0.

References

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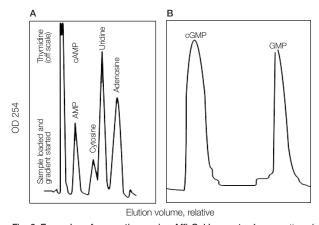


Fig. 2. Examples of separations using Affi-Gel boronate. A, separation of thymidine, cAMP, AMP, adenosine, cytosine, and uridine by a typical gradient elution. A sample containing 0.5 µmol of each molecule was loaded on an equilibrated 0.7 x 45 cm bed of Affi-Gel boronate gel; 0.1 M phosphate was applied after the first peak emerged. The flow rate was 40 ml/hr. B, separation of cGMP and GMP. A sample containing 1.0 µmol cGMP and GMP was loaded onto a 1 x 4 cm column equilibrated with 0.1 M HEPES, pH 8.5. GMP was eluted with 0.1 M NaPO₄, pH 6.0, and detected by a UV monitor.

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Ordering Information

Catalog # Description

153-6103 Affi-Gel Boronate Gel, 5 g 153-6104 Affi-Gel Boronate Gel, 50 g



Bio-Rad Laboratories, Inc.

Life Science Group Web site www.bio-rad.com USA (800) 4BIORAD Australia 02 9914 2800 Austria (01)-877 89 01 Belgium 09-385 55 11 Brazil 55 21 2527 3454 Canada (905) 712-2771 China (86 21) 6426 0808 Czech Republic + 420 2 41 43 05 32 Denmark 44 52 10 00 Finland 09 804 22 00 France 01 47 95 69 65 Germany 089 318 84-0 Greece 30 210 777 4396 Hong (852) 2789 3300 Hungary 36 1 455 8800 India (91-124)-2398112/3/4, 5018111, 6450092/93 Israel 03 951 4127 Italy 39 02 216091 Japan 03-5811-6270 Korea 82-2-3473-4460 Latin America 305-894-5950 Mexico 55-52-00-05-20 The Netherlands 0318-540666 New Zealand 64 9 415 2280 Norway 23 38 41 30 Poland + 48 22 331 99 99 Portugal 351-21-472-7700 Russia 7 095 721 1404 Singapore 65-64153188 South Africa 00 27 11 4428508 Spain 34 91 590 52 00 Sweden 08 555 12700 Switzerland 061 717 95 55 Tailvan (886 2) 2578 7189/2578 7241 United Kingdom 020 8328 2000